



Received: 22 June, 2021
Accepted: 30 June, 2021
Published: 01 July, 2021

*Corresponding author: Dr. Tatsuomi Matsuoka, Department of Biological Science, Faculty of Science and Technology, Kochi University, Kochi 780-8520, Japan, Tel: +81 888448315; Fax: 8188448356; E-mail: tmatsuok@kochi-u.ac.jp

Keywords: *Anisakis larvae*; Seirogan; Wood creosote

<https://www.peertechzpublications.com>



Research Article

Over-the-counter medicine (Seirogan) containing wood creosote kills *Anisakis larvae*

Kou Matsuoka¹ and Tatsuomi Matsuoka^{2*}

¹Kyowakaibyoin (Kyowakai Hospital), Suita, Osaka 564-0001, Japan

²Department of Biological Science, Faculty of Science and Technology, Kochi University, Kochi 780-8520, Japan

Abstract

Background: *Anisakis* food poisoning is characterized by the onset of severe intestinal and stomach pain caused by eating raw or undercooked seafood that harbors the larvae of an anisakid nematode such as *Anisakis simplex*. Although there is currently no effective drug to kill anisakid nematodes, it has been reported that acetylcholinesterase inhibitors such as the over-the-counter medicine 'Seirogan' strongly suppress the nematode's motility.

Methods: One pill of Seirogan was dissolved in 0.01 M HCl (5 mL or 10 mL), and nematodes were exposed to these test solutions for 30 min. To determine whether the nematodes treated with the Seirogan solutions remained alive or not, the nematodes were exposed to trypan blue solution (widely used for the selective staining of dead tissues).

Results: Most (91.7%) of nematodes whose motility had been prevented by Seirogan treatment (1 pill/10 mL) were stained by trypan blue at 24 h after a 30-min treatment with Seirogan. The majority (83.3%) of Seirogan (1 pill/10 mL)-treated nematodes began to be digested by pepsin treatment within 24 h, whereas all living nematodes not treated with Seirogan remained actively moving despite pepsin treatment. When the posterior 4/5 of a nematode was dipped in Seirogan test solution (1 pill/10 mL), only the dipped portion stopped moving.

Conclusion: A widely available OTC intestinal medicine 'Seirogan' kills *Anisakis* larvae at its normal dose, and the killed nematodes are probably digested in the gastric juice. Seirogan components that are effective for killing nematodes may be absorbed from the nematodes' body surface.

Introduction

Anisakis food poisoning (anisakiasis) is characterized by the onset of severe abdominal pain. It is a parasitic disease caused by eating raw or undercooked seafoods that harbor larvae of the anisakid nematodes such as *Anisakis simplex*, *A. pegreffii* and *Pseudoterranova decipiens* [1,2]. There has been a recent increase in the reported prevalence of anisakiasis worldwide [3,4]. In Japan, anisakiasis accounted for 35% of all cases of food poisoning in 2018 [5]. Once consumed, *Anisakis* larvae attempt to penetrate into the gastrointestinal wall, inducing severe gastrointestinal pain and serious allergic reactions such as anaphylactic shock against allergens (many proteins of unknown function) secreted from the nematodes or substances on the surface of nematodes [2,6-10]. *Anisakis* larvae are most frequently detected in the stomach (gastric anisakiasis),

whereas bowel anisakiasis is not very frequent [1,11]. In gastric anisakiasis, symptoms occur 2-3 h after the consumption of raw fish or other seafoods [12,13]. *Anisakis* larvae are tolerant to gastric acid containing pepsin [14] and apparently many tested compounds including anti-nematodal agents such as ivermectin and albendazole [15]. In cases of gastric anisakiasis, the main effective treatment is an endoscopic removal of nematodes [16], while the effective treatment for intestinal anisakiasis has not yet been established.

An acetylcholinesterase inhibitor was reported to suppress the motility of *Anisakis* larvae (which were considered to be killed) [17]. The major component of the over-the-counter (OTC) medicine 'Seirogan' (Taiko Pharmaceutical Co., Osaka, Japan) is wood creosote, which is an acetylcholinesterase inhibitor [18]. Seirogan has been used for >100 years in Asia

as a treatment for stomachache or diarrhea due to digestive disorders. Ten years ago, it was reported that the administration of Seirogan alleviates gastric anisakiasis symptoms (a report of two cases) [19]. In addition, *in vitro* experiment showed that Seirogan quickly suppressed the motility of *Anisakis* larvae, and resulted in the deformation of the nematodal head and disruption of sheath striation [19]. Based on these results, Seirogan was suggested to suppress the viability of nematodes [19]. However, it is still unclear whether the Seirogan-treated immobile nematodes are killed or temporarily paralysed. The allergens secreted by living nematodes may be responsible for an abrupt hypersensitivity reaction leading to the severe stomach pain or other allergic symptoms [2,9]. If the nematodes are killed by Seirogan administration, the production and secretion of allergens by living nematodes may be stopped, thereby resulting in alleviation of allergic symptom. The nematodes rarely penetrate through the gastrointestinal wall to reach the peritoneal cavity [20]. Such a high-risk case may be avoided if the nematodes are killed by Seirogan administration before they deeply penetrate into the gastrointestinal wall. Therefore, we conducted the present study to determine whether or not Seirogan kills *Anisakis* larvae.

Materials and methods

Samples

Anisakis simplex L3 larvae (15–30mm) were isolated from internal organs of mackerel and pacific cod, and then maintained in 0.9% NaCl at room temperature (~25°C). *A. simplex* larva was identified by the shape of stomach and protrusion of posterior end.

Treatment with seirogan

One pill (containing 44 mg of wood creosote) of Seirogan (Taiko Pharmaceutical Co.) was dissolved in 5 mL or 10 mL of 0.01 M HCl (pH 2). *Anisakis* larvae kept in 0.9% NaCl were transferred into small petri dishes filled with a Seirogan solution or with 0.01 M HCl, and the petri dishes were then kept in an incubator for 30 min at 37°C. After treatment, the nematodes were transferred into 0.9% NaCl and kept at room temperature (~25°C) for 2 days of observation.

Trypan blue staining

To determine whether the *Anisakis* larvae that were treated with the Seirogan test solutions remained alive or not, the nematodes were transferred into 0.4% trypan blue solution (Fujifilm Wako Pure Chemical Co., Osaka, Japan) and left them to be stained for 5 h at 37°C.

Pepsin treatment

After Seirogan treatment (1 pill/10 mL) for 30 min, the nematodes were transferred into a 0.01 M HCl solution (pH 2) containing 13.8 µg/mL pepsin (Fujifilm Wako Pure Chemicals) and kept for 24–48 h at 37°C.

Results and discussion

When *Anisakis* larvae were treated with either of the two

different concentrations of Seirogan solutions, all nematodes stopped moving within 30 min. The treated nematodes were transferred into 0.9% NaCl and then observed at 0, 24 and 48. The motility of the nematodes treated with a Seirogan solution (1 pill/5 mL) was not regained at all, whereas 13.3% of the nematodes treated with a diluted Seirogan solution (1 pill/10 mL) began to move 1 day later (Figure 1). In the 0.01 M HCl solution, all nematodes continued to move for ≥2 days (Figure 1).

If motionless nematodes are killed, they will be stained blue by trypan blue staining, which has been widely used for the selective staining of dead tissues or cells [21]. To determine whether Seirogan-treated immobile nematodes are killed or simply temporarily paralyzed, Seirogan (1 pill/10 mL)-treated nematodes were transferred into 0.9% NaCl and kept for 24 h at room temperature, and only immobile nematodes were transferred into 0.4% trypan blue solution. The living nematodes without Seirogan treatment were not stained at all (Figure 2A), whereas the Seirogan-treated immobile nematodes were stained blue (Figure 2B). However, it was difficult to stain the nematodes immediately after a 30-min treatment with Seirogan solution (photographs not shown). In that case, the tissues of nematodes just after Seirogan treatment may have been alive. The results of the trypan blue staining are summarized in Table 1: Most (91.7%) of the Seirogan-treated (1 pill/10 mL) immobile nematodes were stained blue at 24 h after a 30-min treatment with Seirogan. This result suggests that most of the immobile nematodes may be killed within 24 h, but some of the immobile nematodes may be alive.

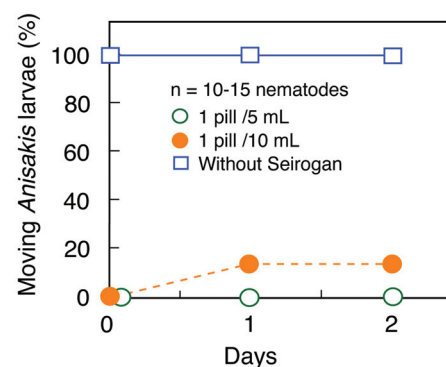


Figure 1: Nematocidal effect of Seirogan on *Anisakis* larvae. Nematodes were treated with Seirogan solution (1 pill/5 mL, open circles), diluted one (1 pill/10 mL, closed circles), and 0.01 M HCl (without Seirogan) (open squares).

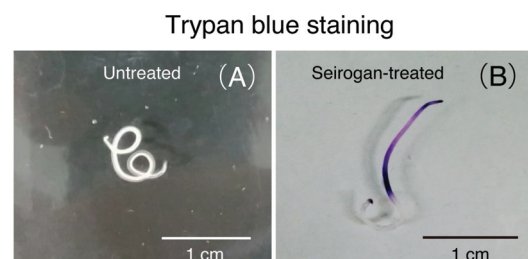


Figure 2: Trypan blue staining. (A) A living nematode without Seirogan treatment. (B) An immobile nematode at 1 day after Seirogan treatment (1 pill/10 mL).

Living *Anisakis* larvae are resistant to human gastric fluid, which contains the digestive enzyme pepsin [14]. The question thus arises: Are the nematodes killed by Seirogan treatment digested by pepsin? The estimated concentration of pepsin contained in human gastric fluid is 13.8 µg/mL [22]. When the living nematodes without Seirogan treatment were incubated in a pepsin solution for 24 h at 37°C, they were still moving actively (Figure 3A). On the other hand, the nematodes treated with Seirogan solution (1 pill/10 mL) were disrupted by pepsin digestion (Figure 3B, arrowheads). The results of the pepsin treatment were summarized in Table 2, showing that most (83.3%) of Seirogan-treated (1 pill/10 mL) immobile nematodes were digested by pepsin treatment. These results suggest that *Anisakis* larvae may begin to be digested within 24 h in the stomach cavity only if they are first killed by oral administration of Seirogan.

When the 4/5 portion from the posterior end of the nematode body was dipped in Seirogan solution (1 pill/5 mL) at room temperature, the posterior 4/5 portion stopped moving soon. The anterior 1/5 portion continued to actively move during observation for 10 min (8 nematodes observed). This result suggests that Seirogan components that are effective for killing nematodes may be absorbed from the nematodes' body surface, and that *Anisakis* larvae can be killed by Seirogan administration even if they partially penetrated into the gastric wall.

Table 1: Trypan blue staining of Seirogan-treated *Anisakis* larvae.

	Stained nematodes (total number)	Rate of stained nematodes (%)
Without treatment	0 (9)	0
Just after treatment	0 (10)	0
24 h after treatment	11 (12)	91.7
48 h after treatment	9 (9)	100

Pepsin digestion

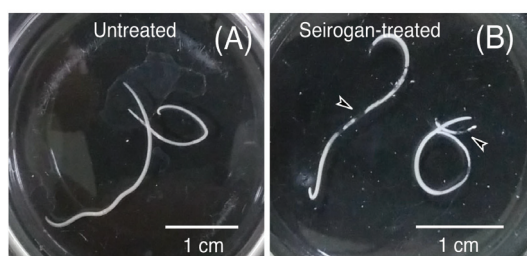


Figure 3: Pepsin digestion for 24 h. (A) Living nematodes without Seirogan treatment. (B) Immobile nematodes just after a 30-min Seirogan treatment (1 pill/10 mL).

Table 2: Pepsin treatment of *Anisakis* larvae.

	Digested nematodes (total number)	Rate of digested nematodes (%)
Seirogan-untreated	0 (10)	0
Seirogan-treated, pepsin for 24 h	10 (12)	83.3
Seirogan-treated, pepsin for 48 h	11 (12)	91.7

The usual dose of Seirogan is three pills (providing 133 mg of wood creosote). The gastric fluid volume in the fasted state of an adult is ~30 mL [23,24]. It is possible that *Anisakis* larvae attached to the stomach wall may be killed by this single dose if three pills of Seirogan are completely dissolved in 30 mL of gastric fluid in the fasted state. If the nematodes are killed by Seirogan before they deeply penetrate into the gastrointestinal wall, a high-risk situation in which the nematodes penetrate through the gastrointestinal wall to reach the peritoneal cavity may be avoided. The secretion of allergens from killed nematodes may also be stopped. The results of the present *in vitro* experiments demonstrate that a widely available OTC intestinal medicine killed *Anisakis* larvae at its normal dose. As the next step, a clinical study should be conducted to elucidate whether the nematodes that have attached to the stomach wall can be actually killed by oral administration of Seirogan.

Conclusion

The present *in vitro* study suggested that *Anisakis* larvae might be killed by oral administration of widely available OTC intestinal medicine 'Seirogan' at its normal dose, and the killed nematodes are probably digested by pepsin contained in the gastric juice. *Anisakis* larvae can be killed by Seirogan even if they partially penetrate into the gastric wall, because Seirogan components may be absorbed from the nematodes' body surface. To our knowledge, this is the first report of the medicine that possesses strong nematocidal activity against *Anisakis* larvae.

Acknowledgment

The *Anisakis* larvae used in these experiment were supplied by Mr. Daijiro Yuki (Marin Biology Laboratory of Kochi University) and by Sourensha Co., Ltd.

Author contributions

TM designed the study and wrote the manuscript. KM contributed to the writing of the manuscript, discussed the results, and contributed to the final manuscript. Authors have accepted responsibility for the entire content of this manuscript and approved its submission.

References

- Sakanari JA, McKerrow JH (1989) Anisakiasis. *Clin Microbiol Rev* 2: 278-284. [Link: https://bit.ly/363Llfy](https://bit.ly/363Llfy)
- Aibinu IE, Smooker PM, Lopata AL (2019) *Anisakis* Nematodes in fish and shellfish- from infection to allergies. *Int J Parasitol Parasites Wildl* 9: 384-393. [Link: https://bit.ly/3AnGRP4](https://bit.ly/3AnGRP4)
- Bao M, Pierce GJ, Pascual S, González-Muñoz M, Mattiucci S, et al. (2017) Assessing the risk of an emerging zoonosis of worldwide concern: anisakiasis. *Sci Rep* 7: 43699. [Link: https://go.nature.com/3A8IEZn](https://go.nature.com/3A8IEZn)
- Rahmati AR, Kiani B, Afshari A, Moghaddas E, Williams M, et al. (2020) World-wide prevalence of *Anisakis* larvae in fish and its relationship to human allergic anisakiasis: a systematic review. *Parasitol Res* 119: 3585-3594. [Link: https://bit.ly/2TixkrC](https://bit.ly/2TixkrC)
- Watari T, Tachibana T, Okada A, Nishikawa K, Otsuki K, et al. (2021) A review of food poisoning caused by local food in Japan. *J Gen Fam Med* 22:15-23. [Link: https://bit.ly/3y7hQWq](https://bit.ly/3y7hQWq)



6. Audicana MT, Kennedy MV (2008) Anisakis simplex: from obscure infectious worm to inducer of immune hypersensitivity. *Clin Microbiol Rev* 21: 360-369. [Link: https://bit.ly/2Uhf7es](https://bit.ly/2Uhf7es)
7. Shiomi K (2010) Current knowledge on molecular features of seafood allergens. *Shokuhin Eiseigaku Zasshi* 51: 139-152. [Link: https://bit.ly/3hii3z6](https://bit.ly/3hii3z6)
8. Mattiucci S, Fazii P, Rosa AD, Paoletti M, Megna AS, et al. (2013) Anisakiasis and gastroallergic reactions associated with *Anisakis pegreffii* infection, Italy. *Emerg Infect Dis* 19: 496-499. [Link: https://bit.ly/2UfkeLP](https://bit.ly/2UfkeLP)
9. Villazanakretzer DL, Napolitano PG, Cummings KF, Magann EF (2016) Fish parasites: a growing concern during pregnancy. *Obstet Gynecol Surv* 71: 253-259. [Link: https://bit.ly/3his6nY](https://bit.ly/3his6nY)
10. Ivanović J, Baltić MŽ, Bošković M, Kilibarda N, Dokmanović M, et al. (2017) Anisakis allergy in human. *Trends Food Sci Technol* 59: 25-29. [Link: https://bit.ly/3Af5r4D](https://bit.ly/3Af5r4D)
11. Yasunaga H, Horiguchi H, Kuwabara K, Hashimoto H, Matsuda S (2010) Short report : Clinical features of bowel Anisakiasis in Japan. *Am J Trop Med Hyg* 83: 104–105. [Link: https://bit.ly/3x6s4G9](https://bit.ly/3x6s4G9)
12. Daschner A, Pascual CY (2005) *Anisakis simplex*: sensitization and clinical allergy. *Curr Opin Allergy Clin Immunol* 5: 281-285. [Link: https://bit.ly/2UU0Gx5](https://bit.ly/2UU0Gx5)
13. Ugenti I, Lattarulo S, Ferrarese F, De Ceglie A, Manta R, et al. (2007) Acute gastric anisakiasis: an Italian experience. *Minerva Chir* 62: 51-60. [Link: https://bit.ly/3drEX6b](https://bit.ly/3drEX6b)
14. Jeon CH, Kim JH (2015) Pathogenic potential of two sibling species, *Anisakis simplex* (s.s.) and *Anisakis pegreffii* (Nematoda: Anisakidae): In vitro and in vivo studies. *Biomed Res Int* 2015: 983656. [Link: https://bit.ly/3jqliWT](https://bit.ly/3jqliWT)
15. Dziekońska-Rynko J, Rokicki J, Jablonowski Z (2002) Effects of ivermectin and albendazole against *Anisakis simplex* in vitro and in guinea pigs. *J Parasitol* 88: 395-398. [Link: https://bit.ly/362TC3y](https://bit.ly/362TC3y)
16. Pravettoni V, Primavesi L, Piantanida M (2012) *Anisakis simplex*: current knowledge. *Eur Ann Allergy Clin Immunol* 44: 150-156. [Link: https://bit.ly/3ds18cl](https://bit.ly/3ds18cl)
17. Víctor López V, Cascella M, Benelli G, Maggi F, Gómez-Rincón C (2018) Green drugs in the fight against *Anisakis simplex*—larvicidal activity and acetylcholinesterase inhibition of *Origanum compactum* essential oil. *Parasitol Res* 117: 861-867. [Link: https://bit.ly/3jt9jZO](https://bit.ly/3jt9jZO)
18. Ogata N, Tagishi H, Tsuji M (2020) Inhibition of acetylcholinesterase by wood creosote and simple phenolic compounds. *Chem Pharm Bull* 68: 1193-1200. [Link: https://bit.ly/2SCRDzV](https://bit.ly/2SCRDzV)
19. Sekimoto M, Nagano H, Fujiwara Y, Watanabe T, Katsu K, et al. (2011) Two cases of gastric Anisakiasis for which oral administration of a medicine containing wood creosote (Seirogan) was effective. *Hepatogastroenterology* 58: 1252-1254. [Link: https://bit.ly/3Abydmw](https://bit.ly/3Abydmw)
20. Furukawa A (1974) Anisakiasis- Its history and a case of acute ileitis, attributable to a living larva of *Anisakis* in the peritoneal cavity. *J Jpn Surg Assoc* 35: 63-69. [Link: https://bit.ly/2SHcKRG](https://bit.ly/2SHcKRG)
21. Mascotti K, McCullough J, Burger S (2000) HPC viability measurement: trypan blue versus acridine orange and propidium iodide. *Transfusion* 40: 693-696. [Link: https://bit.ly/363MYds](https://bit.ly/363MYds)
22. Foltz E, Azad S, Everett ML, Holzknecht ZE, Sanders NL, et al. (2015) An assessment of human gastric fluid composition as a function of PPI usage. *Physiol Rep* 3: e12269. [Link: https://bit.ly/3y3Wrxh](https://bit.ly/3y3Wrxh)
23. Lydon A, Murray C, McGinley J, Plant R, Duggan F, et al. (1999) Cisapride does not alter gastric volume or pH in patients undergoing ambulatory surgery. *Can J Anaesth* 46: 1181-1184. [Link: https://bit.ly/3w9C14q](https://bit.ly/3w9C14q)
24. Mudie DM, Murray K, Hoad CL, Pritchard SE, Garnett MC, et al. (2014) Quantification of gastrointestinal liquid volumes and distribution following a 240 mL dose of water in the fasted state. *Mol Pharm* 11: 3039-3047. [Link: https://bit.ly/7Qg8](https://bit.ly/7Qg8)

Discover a bigger Impact and Visibility of your article publication with Peertechz Publications

Highlights

- ❖ Signatory publisher of ORCID
- ❖ Signatory Publisher of DORA (San Francisco Declaration on Research Assessment)
- ❖ Articles archived in worlds' renowned service providers such as Portico, CNKI, AGRIS, TDNet, Base (Bielefeld University Library), CrossRef, Scilit, J-Gate etc.
- ❖ Journals indexed in ICMJE, SHERPA/ROMEO, Google Scholar etc.
- ❖ OAI-PMH (Open Archives Initiative Protocol for Metadata Harvesting)
- ❖ Dedicated Editorial Board for every journal
- ❖ Accurate and rapid peer-review process
- ❖ Increased citations of published articles through promotions
- ❖ Reduced timeline for article publication

Submit your articles and experience a new surge in publication services (<https://www.peertechz.com/submission>).

Peertechz journals wishes everlasting success in your every endeavours.

Copyright: © 2021 Matsuoka K, et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.